No.

201500325

THE UNITED STATES OF AMERICA

TO ALL TO WHOM THESE PRESENTS SHALL COME:

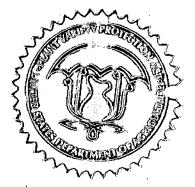
University of Idaho

Whereas, there has been presented to the

Secretary of Agriculture

An application requesting a certificate of protection for an alleged distinct variety of sexually reproduced, or tuber propagated plant, the name and description of which are contained in the application and exhibits, a copy of which is hereunto annexed and made a part hereof, and the various requirements of law in such cases made and provided have been complied with, and the title thereto is, from the records of the PLANT VARIETY PROTECTION OFFICE, in the applicant(s) indicated in the said copy, and whereas, upon due examination made, the said applicant(s) is (are) adjudged to be entitled to a certificate of plant variety protection under the law.

Now, therefore, this certificate of plant variety protection is to grant unto the said applicant(s) and the successors, heirs or assigns of the said applicant(s) for the term of TWENTY years from the date of this grant, subject to the payment of the required fees and periodic replenishment of viable basic seed of the variety in a public repository as provided by law, the right to exclude others from selling the variety, or offering it for sale, or reproducing it, or importing it, or exporting it, or conditioning it for propagation, or stocking it for any of the above purposes, or using it in producing a hybrid or different variety there from, to the extent provided by the PLANT VARIETY PROTECTION ACT. (84 STAT. 1542, AS AMENDED, 7 U.S.C. 2321 ET SEQ.)



Attest:

POTATO

'Northwest Norkotah 90'

In Testimony Whereof, I have hereunto set my hand and caused the seal of the Plant Variety Protection Office to be affixed at the City of Washington, D.C. this third day of June, in the year two thousand and sixteen.

Commissioner

Plant Variety Protection Office Agricultural Marketing Service Cleur J. Vilsul
Secretary of Agriculture

REPRODUCE LOCALLY. Include form number and date	on all reproductions	Form Approved - OMB No. 0581-0055						
AGRICULTURAL MARKE	TING SERVICE	The following statements are made in accordance with the Privacy Act of 1974 (5 U.S.C 552a) and the Paperwork Reduction Act (PRA) of 1995. Application is required in order to determine it a plant variety protection certificate is to be issued						
APPLICATION FOR PLANT VARIETY (Instructions and information collection	PROTECTION CERTIFICATE burden statement on reverse)	(7 U.S.C. 242	Information is hold confidential until cent	tificate is iss	ued (7 U.S.C. 2426).			
1. NAME OF OWNER	RESS (Street and No., or R.F.D. No., City, State, and ZIP Code, and Country) e of Technology Transfer ill Hall PO Box 443003 DB 83844 -3003 HE OWNER NAMED IS NOT A "PERSON", CIVE OF ORGANIZATION (corporation, partnership, lion, etc.) d Grant University Not for Profit d Grant University Not for Profit d Grant University Not for Profit Hall PO Box 443003 SCOW, ID 8385 EPHONE (include area code) B 885-4550 or 529-8376 DP KIND (Common Name) AND SPECIES NAME OF CROP INUS AND SPECIES		RY DESIGNATION OR EXPERIMENTAL I	Control of the control				
University of Idaho		IDRN	90, NWN 90	N	lorthwest Norkotah 90			
4. ADDRESS (Street and No., or R.F.D. No., City, St. Office of Technology Transfer	ate, and ZIP Code, and Country)		NE (include area code) 35-4550	PVP	FOR OFFICIAL USE ONLY O NUMBER			
Morrill Hall PO Box 443003		6. FAX (inclu	de area code)	7	201500325			
Moscow ID	83844 -3003	208-88	5-6127	FILI	NG DATE			
7 IF THE OWNER NAMED IS NOT A "PERSON", GI FORM OF ORGANIZATION (corporation, partnership, association, etc.)		9 DATE OF	NCORPORATION		5/11/15			
Land Grant University Not for Pr	ofit Idaho	1947						
Karen Stevenson and Jeffrey C Office of Technology Transfer Morrill Hall PO Box 443003	. Stark		n listed will receive all papers)	FEES RECEIVE	\$ 4382.00 5/11/15 DATE CERTIFICATION FEE: \$ DATE			
Moscow,		4 -30	13. E-MAIL	D				
(208) 885-4550 or 529-8376	(208) 885-4551 or 522-2	954	karens@uidaho.edu or	jstark(Duidaho.edu			
14. CROP KIND (Common Name)			18 DOES THE VARIETY CONTAIN AT	NY TRANSO	SENES? (OPTIONAL)			
Potato	Contribute of State o	TION HYRPID?	YES NO IF SO, PLEASE GIVE THE ASSIGNED	USDA-API	HIS REFERENCE NUMBER FOR THE			
Solanum tuberosum		TIONTITIBRIDA	APPROVED PETITION TO DEREGULA COMMERCIALIZATION.	TE THE GI	ENETICALLY MODIFIED PLANT FOR			
(Follow instructions on reverse) a	If the Variety If y If anety (Optional) Owner's Ownership If seeds or, for tuber propagated varieties, we maintained in an approved public repositor, de payable to "Treasurer of the United on Office)		OF CERTIFIED SEED? (See Section of Certified Seed?) YES (if "yes", answer item NO (if "no", go to item 23) UNDECIDED 21. DOES THE OWNER SPECIFY THE NUMBER OF CLASSES? YES NO IF YES, WHICH CLASSES? YES NO IF YES, WHICH CLASSES? YES NO IF YES, SPECIFY THE NUMBER 1 FOUNDATION REGIST (If additional explanation is necessary) THE STATE VARIETY OR ANY COMPRESSED.	S 21 and 22 AT SEED O FOUNDATION AT SEED O 2.3, atc. F FERED T AT SEED O	F THIS VARIETY BE LIMITED AS TO ON REGISTERED CERTIFIED F THIS VARIETY BE LIMITED AS TO OR EACH CLASS. CERTIFIED USE the space indicated on the reverse.) THE VARIETY PROTECTED BY			
OTHER COUNTRIES? YES NO IF YES, YOU MUST PROVIDE THE DATE OF F	IRST SALE, DISPOSITION, TRANSFER, (OR USE	YES NO	DATE OF F	BREEDER'S RIGHT OR PATENT)? FILING OR ISSUANCE AND ASSIGNED			
for a tuber propagated variety a tissue culture wi The undersigned owner(s) is(are) the owner of the entitled to protection under the provisions of Section 4	If be deposited in a public repository and n is sexually reproduced or tuber propagated 2 of the Plant Variety Protection Act	naintained for the diplant variety, an sult in penalties	duration of the certificate d believe(s) that the variety is new, distinct,					
NAME (Proof phil opyra)		A comment of the contract of t	(Pease print or type) Karen A. Stevenson					
CAPACITY OF TITLE	IDATE			ITE				
Research Professor	7 Mary 20		icensing Associate	1	1 May 2015			

Exhibit A Form

1. Describe the genealogy (back to and including public and commercial varieties, lines, or clones used) and the breeding method(s).

In August of 2002, 300 single-hill selections were made by the University of Idaho Plant Breeding Team, combining 100 selections from each of three commercial fields in southeast Idaho planted to standard Russet Norkotah. Selection of each hill was based on late-season vigor, late maturity, and large vine size. An attempt was made to identify clones fitting one of two categories: those with extreme expression of vigor (giant hills) that were obviously later than the surrounding plants; and those with a more subtle expression (intermediate types) of later maturity and larger size in comparison to surrounding plants. All tubers from the selected hills were harvested, bagged, and stored for planting the following year. In addition to the selections, four random plants expressing typical Russet Norkotah vine type were harvested from each of the 3 fields (for a total of 12) and stored to use as check clones.

Give the details of	Subsequent stages of selection and multiplication.	2.0
Year	Detail of Stage	Selection Criteria
2003	The 300 selected clones along with the 12 check clones were planted for evaluation at the University of Idaho Aberdeen R & E Center. The trial consisted of two replications of 10 hills each. The plots were planted in units of four hills and were aggressively rouged for PVY.	During the growing season, assessment was made of vine size, vine maturity, and early die symptoms. At season's end, the plots were harvest and tubers weighed and graded to determine yield and value. Of the 300 initial clones, 82 were selected for further evaluation based on greater vine vigor, disease resistance and yield.
2004	Tubers of the selected lines were indexed for PVY and a small seed increased of disease free material was established and maintained by USDA/ARS cooperators (specifically Jonathan Whitworth). Using other tubers left over from the 2003 trial, a four replicate (20 hills each) trial of remaining clones was planted at the Aberdeen R & E Center	The process of evaluation describe for 2003 we repeated. Following data assessment, 14 clones were advanced into 2005 evaluations. Of the 14 clones, seven expressed giant hill typ while the other seven were more intermediate expression of vine size and maturity.
2005	Seed derived from the ARS increases were used to plant a four replicate field trial (24 hills each) in Riverton, Idaho and a single replicate trial near Bliss, Idaho.	Based on the data from the two trials, five clone were identified that expressed a combination of high yield, good tuber appearance, and good resistance to early dying symptoms. Two of the clones, including NWN 90 expressed extreme giant hill type, vigor and lateness, and had higher yield than the check clones.
2006-2011	Seed of the five clones was increased by several grower cooperators and the clones were evaluated in multiple field trials each year across southern Idaho.	The clones again were evaluated for high yield tuber quality and appearance, and resistance to early dying.
2012-2014	Two of the clones, including NWN 90, were selected as having the greatest potential for commercial production and were evaluated in replicated field trials in Rupert, Parma, Kimberly, Aberdeen and Rexburg, Idaho.	Evaluations were conducted on the two clones in comparison with standard Russet Norkotah obtained from seed growers. In addition to yie and quality, a detailed analysis of seasonal vin and tuber growth, vine senescence, disease resistance and biochemical properties of the tubers were performed.

3a. Is the variety uniform? \underline{X} Yes No

How did you test for uniformity?

Northwest Norkotah 90 has been clonally propagated since the first year of selection. The variety has remained uniform during all of the subsequent years of maintenance and propagation.

3b.Is the variety stable?X YesNo

How did you test for stability? Over how many generations?

Northwest Norkotah 90 has been clonally propagated for 13 years of evaluations. It has shown stability over 13 generations and has not produced any recognizable variants.

4. Are genetic variants observed or expected during reproduction and multiplication? _____ Yes __X_No

If yes, state how these variants may be identified, their type and frequency.

Exhibit B Form

Based on overall morphology, <u>'Northwest Norkotah 90'</u> is most similar to <u>'Russet Norkotah'</u>

Applicant's new variety is most similar comparison variety(ies)

<u>'Northwest Norkotah 90'</u> most clearly differs from <u>'Russet Norkotah'</u> in the following traits:

Applicant's new variety

Most similar comparison variety(ies)

Name the specific trait, and then list the value of that trait for each variety in the comparison. Attach appropriate supporting evidence (see the Guidelines for Presenting Evidence in Support of Variety Distinctness, available from the PVP Office or website).

	Applicant's New Variety 'Northwest Norkotah 90'	1st Comparison Variety 'Russet Norkotah'	Location of Evidence	
1. Qualitative traits:				
Full Flowering	~65 Days after planting	~50 Days after planting	Photographs	
Inflorescence/plant Florets/inflorescence	Medium (2.86) Medium(9.82)	Few (1.47) Few (6.20) aborts	Exhibit C Table 1 - Exhibit D	
Plant Height	Medium tall	Short	Table 2a & 2b	
2. Color traits: * measure	d using the Royal Horticultura	Society Colour Chart (RH	S)	
tuber skin color*	grayed-orange	grayed-orange	Exhibit C and	
	(RHS 165 B)	(RHS 165 C)	photographs	
3. Quantitative traits:				
Protein	Medium (5.04%) at Aberdeen, (5.27%) at Rexburg, Idaho	Low (4.59%) at Aberdeen, (5.08%) at Rexburg, Idaho	Table 3	
Sucrose Sugars	Med-high (0.144% FWB) at Aberdeen, (0.156% FWB) at Rexburg, Idaho	Medium (0.095% FWB) at Aberdeen, (0.126% FWB) at Rexburg, Idaho	Table 4 - Exhibit D	
4. Other:				
Plant Vigor	48% at Aberdeen, 64% at	13% at Aberdeen, 5%	Table 5a & 5b	
0-100%; 0= dead, 100% =lush and vigorously	Rexburg, ID (103 days after planting (DAP))	at Rexburg, ID (103 DAP)		
growing				
Green Vine (g/m ²)	87 g/m ² at Aberdeen, 134 g/m ² at Rexburg, ID (88/89 DAP)	17 g/m ² at Aberdeen, 35 g/m ² at Rexburg, ID (88/89 DAP)	Table 6a & 6b – Exhibit D	

FWB= Fresh Weight Basis

Use additional tables to present clear differences for additional comparison varieties. Use additional pages to present supporting evidence

NORTHWEST NORKOTAH 90

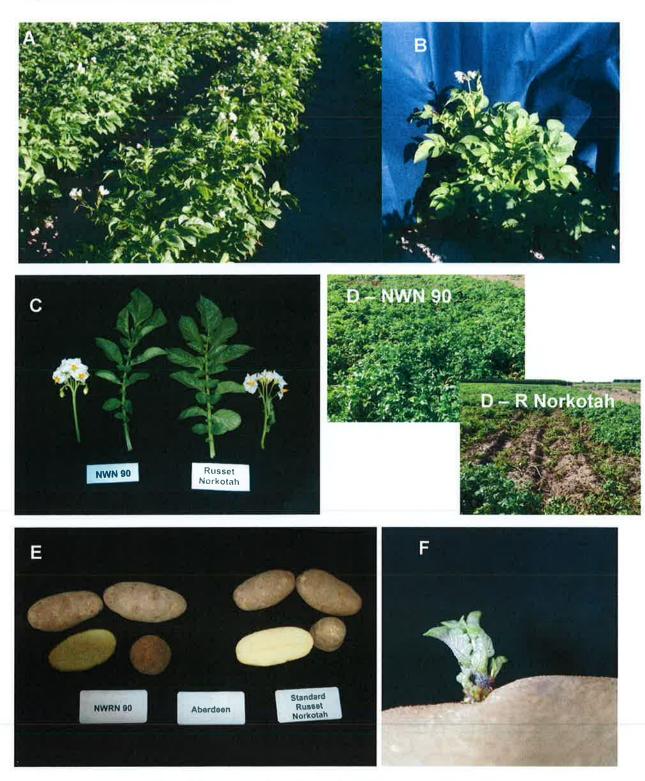
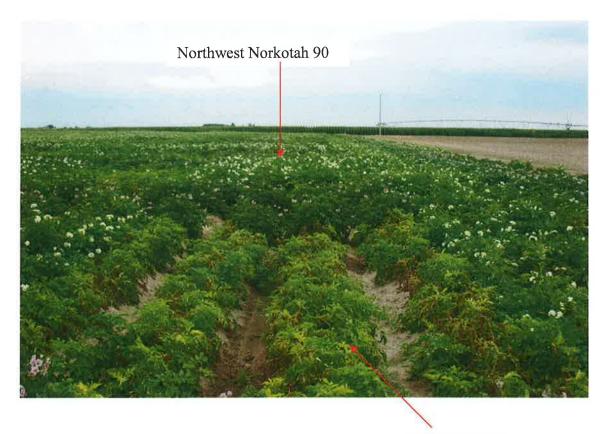


Figure 1. Photographs of NWN 90 showing a) field plants, b) whole plant, c) compound leaf and flower compared to Russet Norkotah, d) vine maturity of NWN 90 and Russet Norkotah on August 15, 2014, e) external tuber appearance and tuber flesh color, and f) light sprout.



Russet Norkotah

Figure 2. Plants in bloom at Aberdeen, Idaho approximately 68 days after planting. Russet Norkotah plants in front and Northwest Norkotah 90 in back.

Northwest Norkotah 90

Russet Norkotah



Plant Senescence August 15, 2014 Kimberly, Idaho



Plant Senescence August 19, 2014 Aberdeen, Idaho



Plant Senescence August 21, 2014 Rexburg, Idaho

Figure 3. Northwest Norkotah 90 (left) plant senescence compared to Russet Norkotah (right) at Kimberly, Aberdeen, and Rexburg, Idaho.

Application for Plant Variety Protection Certificate

Exhibit D: Additional Description Information

Variety: Northwest Norkotah 90

Owner: Idaho Agricultural Experiment Station

Northwest Norkotah 90 (NWN 90) flowers about 65 days after planting (DAP) and Standard Russet Norkotah (RN) flowers earlier at ~ 50 DAP. NWN 90 produces more inflorescence per plant with more florets per inflorescence than Russet Norkotah (Table 1).

At Aberdeen and Rexburg, Idaho data was collected weekly on a 2 plant sample with in each of 4 replications, starting ~75 days after planting. Plant height was measured in cm (Tables 2a and 2b). Plant vigor was determined on a scale of 0-100% (Tables 5a and 5b). The entire plant including vines and tubers were collected and vines were separated into green and lush vines, yellow clorotic vines, and dead vines. Fresh weight of each vine category and tubers were measured and samples were dried and reweighed. The total amount of green vines grams per meter² were then calculated (Tables 6a and 6b).

In direct comparison with Russet Norkotah, NWN 90 tubers have higher protein content (5.2% protein for Northwest Norkotah 90 vs. 4.8% protein for Russet Norkotah averaged over 2 locations) Table 3. Mean protein content for individual years were 5.04% for NWN 90 and 4.59% for Russet Norkotah at Aberdeen, Idaho (p=0.05) and 5.27% for NWN 90 and 5.08% for RN at Rexburg, Idaho (p=0.05).

Two location average sucrose sugar contents were 0.150% FWB for Norwest Norkotah 90 vs. 0.105% FWB for Russet Norkotah Table 4. Mean total sucrose sugar contents for individual locations were 0.144% FWB for NWN 90 and 0.095% FWB for Russet Norkotah at Aberdeen, (p=0.05) and 0.156% FWB for NWN 90 and 0.126% FWB for RN at Rexburg, Idaho (p=0.05).

Protocols are attached. Statistical analysis was performed using the GLM procedure within SAS.

Protocols used for comparisons between Northwest Norkotah 90 and Russet Norkotah.

Protocol used for Northwest 90 and Russet Norkotah comparisons

Variety Yield trials were conducted using a randomized complete block design of 20' plots using 4 replications. Trials were planted on May 1, 2014 at Aberdeen, Idaho, and April 24 at Kimberly, Idaho, and May 15 at Rexburg, Idaho.

At Aberdeen and Rexburg, Idaho data was collected weekly on a 2 plant sample within each replication, starting ~75 days after planting. Plant height was measured in cm. Plant vigor was determined on a scale of 0-100%. Plants were collected vines were separated into green and lush vines, yellow clorotic vines, dead vines, and tubers. Fresh weight was measured and samples were dried and reweighed. Green vines grams per meter² were then calculated.

Protocol used for inflorescence and florets

Plants were grown at Aberdeen, Idaho in variety yield trials. Counts were taken on 80 plants in full bloom in early July at about 60 days after planting (DAP). Inflorescence per plant and florets per inflorescence counted.

Protocol used for chemical composition

Tubers were harvested from trials in late September (~140-145 DAP). A five tuber sample from 4 replications was freeze dried and ground to use in assays. Standard Operating Procedures are attached for Protein and Sucrose.

Statistical analysis was preformed using SAS proc GLM.

Table 1. Northwest Norkotah 90 and Russet Norkotah Comparisons for Inflorescence per plant and Florets per inflorescence using the GLM Procedure for Potatoes grown at Aberdeen, Idaho in 2014.

Anova		Inflore	scence	Floi	ets	
Source	DF	F Value	PR > F	F Value	PR > F	
Variety	1	20.46	<.0001	21.97	<.0001	
Replication	72	0.53	0.9964	0.59	0.9859	

Variety		Inflorescence	Florets
Northwest Norkotah 90	Mean	2.86	9.82
	Minimum	0	0
	Maximum	8	18
	Stdev	1.847	2.621
Russet Norkotah	Mean	1.47	6.20
	Minimum	0	0
	Maximum	5	15
	Stdev	1.149	4.224
LSD =0.05		0.616	1.537

Counts were collected on 73 plants grown at Aberdeen, Idaho in 2014.

1.29

21.5

1.91

NS

Table 2a. Northwest Norkotah 90 and Russet Norkotah Comparisons for plant height over time using the GLM Procedure for Potatoes grown at Aberdeen, Idaho in 2014.

				D	ays A	fter P	lantin	g Plan	t Heig	ht (cm	1)			
Anova	Anova		4	83		8	88		103		111		116	
Source	DF	F Value	PR >	F Value	PR >	F Value	PR >	F Value	PR >	F Value	PR >	F Value	PR >	
Variety	1	13.4	0.035	21.9	0.019	10.2	0.049	93.7	0.002	29.9	0.012	4.62	0.120	
Rep	3	0.08	0.969	0.81	0.568	1.70	0.337	1.92	0.303	1.07	0.479	0.40	0.764	
Variet	У													
NWN 90														
Mean		49	0.0	65	65.0		65.3		64.5		41.5		13.7	
Minimu	m	4	6	5	0	5	59		2	2	.5	1	5	
Maximu	ım	5	4	7	4	70		72		5	7	1	.8	
Stdev		3.4	46	10	.89	5.	19	8.66		13	3.7	6.	13	
Russet														
Norkotah	1													
Mean		35	35.3		35.0		41.5		20.3		4.5		5.5	
Minimu	m	2	9	28		30		15		3		4		
Maximu	ım	3	8	4	1	6	66		30		5	8	8	

16.52

23.7

6.85

14.5

Stdev

LSD =0.05

4.27

11.9

5.48

20.4

6.27

18.92

6.24

18.78

Table 2b. Northwest Norkotah 90 and Russet Norkotah Comparisons for plant height over time using the GLM Procedure for Potatoes grown at Rexburg, Idaho in 2014.

			D	ays A	fter P	antin	g Plan	t Heig	ht (cn	1)			
	7	5	83		89		96		103		110		
)F	F Value	PR >	F Value	PR >	F Value	PR >	F Value	PR > F	F Value	PR > F	F Value	PR >	
1	13.9	0.065	4.74	0.161	49.1	0.019	22.19	0.042	64.0	0.015	69.9	0.014	
3	0.37	0.787	0.20	0.889	0.99	0.537	0.79	0.602	1.27	0.469	1.02	0.529	
	64	1.3	66.3		80.7		74.2		76.3		70.0		
	5	8	5	0	7	1	6	6	7	0	6	5	
	7	3	8	2	9	0	8	3	8	32	8	0	
	6.	70	13	.94	7.	76	7.	37	5.	68	6.	45	
	43	3.3	39	9.8	43	3.3	40).4	41	1.8	33	3.0	
	3	9	2	.9	3	8	3	0	3	35	2	5	
	4	.7	47		4	48		51		50		40	
	_	F Value 1 13.9 3 0.37 64 5 7 6.	OF Value F 1 13.9 0.065	75 8 F F PR > F Value 1 13.9 0.065 4.74 3 0.37 0.787 0.20 64.3 66 58 5 73 8 6.70 13 43.3 39 39	75 83 PF F PR > F PR > F PR > 1 13.9 0.065 4.74 0.161 3 0.37 0.787 0.20 0.889 64.3 66.3 58 50 73 82 6.70 13.94	75 83 8 PF F PR > F PR > F Value F Value 1 13.9 0.065 4.74 0.161 49.1 3 0.37 0.787 0.20 0.889 0.99 64.3 66.3 80 58 50 7 73 82 9 6.70 13.94 7. 43.3 39.8 43 39 29 3	75 83 89 OF Value F Value	75 83 89 9 PF Value F Value F Value F Value F Value F Value 1 13.9 0.065 4.74 0.161 49.1 0.019 22.19 3 0.37 0.787 0.20 0.889 0.99 0.537 0.79 64.3 58 50 71 6 73 82 90 8 77 73 82 90 7.76 7.76 43.3 39 29 38 39 39 29 38 33 39	775 83 89 96 OF Value F Value PR > Value <td>75 83 89 96 10 OF Value F Value</td> <td>OF Value F Value F Value PR > F Value F Value PR > F Value PR > Value PR > Value F Value PR > V</td> <td>75 83 89 96 103 113 of Value F Value F Value PR Value F Value PR Value F Value PR Value F Value PR Value F Value</td>	75 83 89 96 10 OF Value F Value	OF Value F Value F Value PR > F Value F Value PR > F Value PR > Value PR > Value F Value PR > V	75 83 89 96 103 113 of Value F Value F Value PR Value F Value PR Value F Value PR Value F Value PR Value F Value	

4.11

22.41

7.72

NS

3.50

23.39

Stdev

LSD =0.05

9.06

12.53

Table 3. Norwest Norkotah 90 and Russet Norkotah Comparisons for Percent Protein using the GLM Procedure for Potatoes grown at Aberdeen, and Rexburg, Idaho in 2014.

Anova		Aberdeen Pe	rcent Protein	Rexburg Percent Protein			
Source	DF	F Value	PR > F	F Value	PR > F		
Variety	1	46.87	<0.0001	8.38	0.0146		
Replication	11	7.34	0.0013	5.85	0.0034		

	Aberdeen Protein (%)	Rexburg Protein (%)
Mean	5.04	5.27
Minimum	4.53	4.86
Maximum	5.58	5.79
Stdev	0.31	0.20
Mean	4.59	5.08
Minimum	4.07	4.59
Maximum	5.11	5.74
Stdev	0.26	0.32
	0.146	0.141
	Minimum Maximum Stdev Mean Minimum Maximum	(%) Mean 5.04 Minimum 4.53 Maximum 5.58 Stdev 0.31 Mean 4.59 Minimum 4.07 Maximum 5.11 Stdev 0.26

Table 4. Norwest Norkotah 90 and Russet Norkotah Comparisons for Percent Sucrose using the GLM Procedure for Potatoes grown at Aberdeen, and Rexburg, Idaho in 2014.

Anova		Aberdeen Per	cent Sucrose	Rexburg Percent Sucrose			
Source	DF	F Value	PR > F	F Value	PR > F		
Variety	1	44.12	0.0003	6.02	0.0439		
Replication	7	0.90	0.5527	0.87	0.5685		

Variety		Aberdeen Sucrose (% Fresh weight basis)	Rexburg Sucrose (% Fresh weight basis)
Northwest 90	Mean	0.144	0.156
	Minimum	0.111	0.118
	Maximum	0.162	0.227
	Stdev	0.017	0.031
Russet Norkotah	Mean	0.095	0.126
	Minimum	0.086	0.106
	Maximum	0.117	0.145
	Stdev	0.009	0.014
LSD =0.05		0.017	0.029

Table 5a. Northwest Norkotah 90 and Russet Norkotah Comparisons for plant vigor over time using the GLM Procedure for Potatoes grown at Aberdeen, Idaho in 2014.

Days After Planting Plant Vigor (0-100%; 0=dead, 100=lush and vigorously growing)

Anov	a	7	4	8	3	8	8	10	03	11	L1	13	L6
Source	DF	F Value	PR >	F Value	PR > F	F Value	PR >	F Value	PR >	F Value	PR > F	F Value	PR >
Variety	1	4.76	0.117	625.0	0.000	4.84	0.115	29.4	0.012	147.0	0.001	121.0	0.002
Rep	3	0.22	0.880	9.00	0.052	0.36	0.788	0.40	0.764	1.00	0.500	1.00	0.500

Variety

97.5	88.8	77.5	47.5	35.0	27.5
95	85	70	40	30	20
100	90	80	60	40	30
2.89	2.50	5.00	9.57	5.77	5.00
86.3	57.5	50.0	12.5	0.0	0.0
80	50	40	10	0	0
95	60	80	20	0	0
7.50	5.00	20.00	5.00	0.00	0.00
NS	3.9	NS	20.5	9.2	7.9
	95 100 2.89 86.3 80 95 7.50	95 85 100 90 2.89 2.50 86.3 57.5 80 50 95 60 7.50 5.00	95 85 70 100 90 80 2.89 2.50 5.00 86.3 57.5 50.0 80 50 40 95 60 80 7.50 5.00 20.00	95 85 70 40 100 90 80 60 2.89 2.50 5.00 9.57 86.3 57.5 50.0 12.5 80 50 40 10 95 60 80 20 7.50 5.00 20.00 5.00	95 85 70 40 30 100 90 80 60 40 2.89 2.50 5.00 9.57 5.77 86.3 57.5 50.0 12.5 0.0 80 50 40 10 0 95 60 80 20 0 7.50 5.00 20.00 5.00 0.00

Table 5b. Northwest Norkotah 90 and Russet Norkotah Comparisons for plant vigor over time using the GLM Procedure for Potatoes grown at Rexburg, Idaho in 2014.

Days After Planting Plant Vigor (0-100%; 0=dead, 100=lush and vigorously growing)

Anov	а	7	5	8	3	8	9	9	6	10	03	13	10
Source	DF	F Value	PR >										
Variety	1	96.0	0.002	243.0	0.001	200.0	0.001	458.4	0.000	129.9	0.001	93.4	0.002
Rep	3	1.00	0.500	1.00	0.500	1.00	0.500	1.00	0.500	0.68	0.618	1.00	0.500

Variety

-						
NWN 90						
Mean	80.0	70.0	67.5	56.3	63.7	36.3
Minimum	80	70	60	50	55	25
Maximum	80	70	75	60	70	40
Stdev	0.00	0.00	6.45	4.79	7.50	7.50
Russet						
Norkotah						
Mean	40.0	25.0	17.5	5.0	5.0	0
Minimum	30	20	15	5	0	0
Maximum	50	30	20	5	10	0
Stdev	8.16	5.77	2.89	0.00	5.77	0.00
LSD =0.05	12.9	9.2	11.3	7.6	16.4	11.9

Table 6a. Northwest Norkotah 90 and Russet Norkotah Comparisons for plant green vine over time using the GLM Procedure for Potatoes grown at Aberdeen, Idaho in 2014.

	Days After Planting Plant Green Vine (g/m²)													
Anova		74		8	83		88		103		111		116	
Source	DF	F	PR >	F	PR >	F	PR >	F	PR >	F	PR >	F	PR >	
		Value	F	Value	F	Value	F	Value	F	Value	F	Value	F	
Variety	1	19.4	0.021	17.7	0.024	106.1	0.002	20.72	0.019	12.9	0.037	1.39	0.328	
Rep	3	0.42	0.754	0.35	0.796	3.06	0.191	1.04	0.487	1.00	0.500	1.00	0.500	
Varie	ty													
NWN 90	-													
Mear	ı	61	5	83.7		86.7		35.4		34.2		10.0		
Minimu	ım	51	6	61	l.1	60	60.9 16.10		.10	6	.2	()	
Maximu	Maximum 73		3.5	110.5		104.9 51.6		L. 6	48	3.3	35	5.5		
Stdev	/	10	.82	22	.30	18	.50	15	5.6	19.05		17	.13	
Russet														
Norkotal	h													
Mear	1	30	.9	22	2.5	17	7.0	0.	22	0	.0	0	.0	
Minimu	ım	29	0.0	15	5.9	10	0.1	0	.0	(כ	()	
Maximu	um	37	. .4	34	34.0		22.8		89	()	()	
Stdev	,	4.	44	8.			39	0.44		0.	00	0.	00	

21.5

24.6

30.3

NS

46.3

22.1

LSD =0.05

Table 6b. Northwest Norkotah 90 and Russet Norkotah Comparisons for plant green vine over time using the GLM Procedure for Potatoes grown at Rexburg, Idaho in 2014.

		Days After Planting Plant Green Vine (g/m²)												
Anova		75		8	83		89		96		103		110	
Source	DF	F Value	PR >	F Value	PR >	F Value	PR >	F Value	PR >	F Value	PR > F	F Value	PR >	
Variety	1	5.15	0.108	14.13	0.033	31.48	0.011	19.73	0.021	36.73	0.009	66.28	0.004	
Rep	3	0.39	0.770	0.72	0.601	1.23	0.434	2.05	0.285	0.46	0.729	1.00	0.500	
NWN 90		00		22	1 6	12	2 6	11	7.6	10	0.6	EG		
Mear		00	3.5	221.6		12	3.6	11	7.6	10	0.6	59	3.2	
Minimu			3.9		3.32	_	2.9		9		5.7		7.2	
Maximu			8.0		5.3		2.2		3.7		5.1		9.2	
Stdev				.03		.26		.96		.30				
Russet														
Norkotał	1													
Mean	1	61	l. 6	41	l.9	34	1.9	16	5.1	15.5		0.0		
Minimu	ım	49	9.2	29	9.5			0		31.9 0 0)	0	

37.6

3.01

56.0

49.2

9.43

152.1

68.9

9.43

NS

Maximum

Stdev

LSD =0.05

32.4

18.61

72.7

32.4

17.95

44.7

0

0.00

22.8

REPRODUCE LOCALLY. Include form number and editi	ion date on all reproductions.					
U.S. DEPARTMENT OF AGRICULTURE AGRICULTURAL MARKETING SERVICE EXHIBIT E STATEMENT OF THE BASIS OF OWNERSHIP	Application is required in order to determine if a plant variety protection certificate is to be issued (7 <i>U.S.C. 2421</i>). The information is held confidential until the certificate is issued (7 <i>U.S.C. 2426</i>).					
1. NAME OF APPLICANT(S)	2. TEMPORARY DESIGNATION	3. VARIETY NAME				
1. WAINE OF AFFEIDAIN (O)	OR EXPERIMENTAL NUMBER	J. VARIETT WANTE				
4. ADDRESS (Street and No., or R.F.D. No., City, State, and ZIP, and Country)	5. TELEPHONE (Include area code)	6. FAX (Include area code)				
	7. PVPO NUMBER	0.1101(maide alea esec)				
8. Does the applicant own all rights to the variety? Mark an "X" in the	e appropriate block. If no, please expla	in. YES NO				
9. Is the applicant a U.S. national or a U.S. based entity? If no, give	name of country. YES	□ NO				
10. Is the applicant the original owner? YES	NO If no, please answer <u>one</u>	of the following:				
	(ana) the eniminal assessor(a) a LLC Netice	-1/-)2				
a. If the original rights to variety were owned by individual(s), is (· · ·	• •				
YES	NO If no, give name of countr	ry				
b. If the original rights to variety were owned by a company(ies)	is (are) the original owner(s) all S ha	sed company?				
YES	NO If no, give name of countr	ту				
11. Additional explanation on ownership (Trace ownership from origin	nal breeder to current owner. Use the n	everse for extra space if needed):				
11. Additional explanation on ownership (Prace ownership from origin	nar broader to darront owner. God the re	everee for extra apade if fleeded).				
DI FASE NOTE:						
PLEASE NOTE:						
PLEASE NOTE: Plant variety protection can only be afforded to the owners (not license)	sees) who meet the following criteria:					
	erson must be a U.S. national, national					
Plant variety protection can only be afforded to the owners (not licens 1. If the rights to the variety are owned by the original breeder, that p	erson must be a U.S. national, national of the U.S. for the same genus and spectured the original breeder(s), the company	ies. y must be U.S. based, owned by				
Plant variety protection can only be afforded to the owners (not license) 1. If the rights to the variety are owned by the original breeder, that penational of a country which affords similar protection to nationals of the variety are owned by the company which employ nationals of a UPOV member country, or owned by nationals of a country or owned by nationals or owned	erson must be a U.S. national, national of the U.S. for the same genus and specture yed the original breeder(s), the company country which affords similar protection	ies. y must be U.S. based, owned by to nationals of the U.S. for the same				
Plant variety protection can only be afforded to the owners (not license) 1. If the rights to the variety are owned by the original breeder, that penational of a country which affords similar protection to nationals of the variety are owned by the company which employ nationals of a UPOV member country, or owned by nationals of a genus and species.	erson must be a U.S. national, national of the U.S. for the same genus and spectyed the original breeder(s), the company country which affords similar protection original owner and the applicant must not the same of the sa	y must be U.S. based, owned by to nationals of the U.S. for the same neet one of the above criteria.				

According to the Paperwork Reduction Act of 1995, an agency may not conduct or sponsor, and a person is not required to respond to a collection of information unless it displays a valid OMB control number. The valid OMB control number for this information collection is 0581-0055. The time required to complete this information collection is estimated to average 0.1 hour per response, including the time for reviewing the instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information.

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> U.S. DEPARTMENT OF AGRICULTURE AGRICULTURAL MARKETING SERVICE SCIENCE AND TECHNOLOGY PLANT VARIETY PROTECTION OFFICE BELTSVILLE, MD 20705

> > **EXHIBIT F**

DECLARATION REGARDING DEPOSIT EMPORARY OR EXPERIMENTAL DESIGNATION ADDRESS (Street and No. or RD No., City, State, and Zip Code and Country) NAME OF OWNER (S) IDRN 90, NWN 90 Morrill Hall 414 University of Idaho PO Box 443003 VARIETY NAME Moscow, Idaho 83844-303 Northwest Norkotah 90 FOR OFFICIAL USE ONLY ADDRESS (Street and No. or RD No., City, State, and Zip Code and Country) NAME OF OWNER REPRESENTATIVE (S) Morrill Hall 414 Karen Stevenson PVPO NUMBER

PO Box 443003

Moscow, Idaho 83844-303

I do hereby declare that during the life of the certificate a viable sample of propagating material of the subject variety will be deposited, and replenished as needed periodically, in a public repository in the United States in accordance with the regulations established by the Plant Variety Protection Office.

Jeffrey C. Stark

1 May 2015

According to the Paperwork Reduction Act of 1995, an agency may not conduct or sponsor, and a person is not required to respond to a collection of information unless it displays a valid OMB control number. The valid OMB control number for this information collection is 0581-0055. The time required to complete this information collection is estimated to average 8.5 hours per response, including the time for review instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information.

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> U.S. DEPARTMENT OF AGRICULTURE AGRICULTURAL MARKETING SERVICE SCIENCE AND TECHNOLOGY PLANT VARIETY PROTECTION OFFICE BELTSVILLE, MD 20705

OBJECTIVE DESCRIPTION OF VARIETY Potato (Solanum tuberosum L.)

INSTRUCTIONS

The Objective Description Form:

The objective description form lists characteristics to be used as the basis for developing the description of potato varieties. It is designed to guide the applicant in describing a variety in detail so a meaningful comparison with other potato varieties can be accomplished. It is recommended that this form be completed in as much detail as possible to ensure an accurate description. Please fill in the requested data and place the appropriate number that describes the varietal characters typical of this potato variety and the reference varieties in the respective boxes.

Test Guidelines:

Any statistical and trial (field test) data that may be necessary to support the variety description should be attached to this form. Please include for trial data the plot size, number of replications, number of plants, plant spacing, trial locations and growing periods. Trials should normally be conducted at one place, in the region that the variety has been adapted for, with a minimum of one growing period in the United States. All comparative data should be determined from varieties entered in the same trials. The size of the plots should be such that plants or parts of plants may be removed for measuring and counting without prejudice to the observations which must be made at the end of the growing period. As a minimum, each test should include a total of 60 plants which should be divided between two or more replicates. Separate plots for observation and measuring can only be used if they have been subject to similar environmental conditions. To determine color for a plant or plant parts a recognized standard color chart must be used such as the Royal Horticultural Society (RHS) Color Chart or Munsell Color Chart (MCC).

Reference Varieties:

The application variety should be compared to at least one reference variety preferably a set of reference varieties. The reference varieties should be market class standard varieties currently grown in the United States and or the variety (ies) most similar. The following varieties are recommended as market class standards to be used as reference varieties:

Round-white table-stock	Superior
Chip-processing	Atlantic, Snowden, Norchip
Frozen-processing	Russet Burbank
Russet table-stock	Russet Burbank, Russet Norkotah, Goldrush
Red table-stock	Red Pontiac, Red Norland, Red Lasoda

If the applicant does not use one of the recommended reference varieties by the PVP office, a complete description of the reference variety should be submitted by the applicant (Exhibit C).

Characteristics:

Light sprout characteristics are supplied in **Figure 1**. The plant type and growth habit characteristics are collected at early first bloom. **Figure 2** is supplied to help visualize the growth habit. For this descriptor, look at the stems rather than the stems and foliage. Plant maturity is measured at natural vine senescence.

Stem characteristics are also collected at early bloom. Stem anthocyanin coloration is divided into two descriptors: Location and intensity. **Figure 3** is supplied to give an example of stem wings.

Leaf characteristics are observed at early first bloom. Fully-developed leaves located on the middle third of the plant should be used. Leaf pubescence refers to general trichomes. **Figure 4** is supplied for examples of leaf silhouette. Leaf stipules are shown in **Figure 5** for visual definition. **Figure 6** is supplied to define leaf characteristics. **Figure 7** should be used to describe terminal and primary leaflet shape. **Figures 8 and 9** are used to describe the terminal and primary leaflet shape of tip and base, respectively. To measure the total number of primary leaflets pairs, collect 10 fully developed petioles (with leaves attached from each replication) and take the average number of secondary and tertiary leaflets. Glandular trichomes should be described in the Additional Comments and Characteristics (Descriptor 15).

Inflorescence characteristics should be measured at early first bloom. **Figures 10, 11 and 12** are supplied to describe anther and stigma shape, respectively. Corolla, calyx, anther, stigma, and pollen should be observed on newly opened flowers. Berry production should be based on field-grown plants rather than greenhouse plants.

Tuber characteristics should be observed following harvest. **Figures 13 and 14** are available to describe distribution of secondary color and tuber shape, respectively.

Disease and pest reactions should be based upon specific tests or statistical analysis rather than just field observations, rating 1 as Highly Resistance and 9 as Highly Susceptible, please follow the scale on each descriptor. Other diseases or pests reactions not requested can be described if it is felt that it would be helpful to determine novelty of the variety.

Quality characteristics should be described according to the market use.

If the plant is transgenic, this gene insertion(s) should be described.

Chemical identification and any other characteristics can be described if they are helpful in distinguishing the variety.

Legend:

V = Application Variety

R1-R4 = Reference Varieties

* = Both the reference variety (ies) and application variety must be described for characteristics designated with an asterisk.

2. LIGHT SPROUT CHARACTERISTICS: (continued)

LIGHT SPROUT TIP: PUBESCENCE

1 = Absent

2 = Weak

3 = Medium

4 = Strong

5 = Very Strong

R1

R2

R3

R4

LIGHT SPROUT TIP ANTHOCYANIN COLORATION

2 = Red-violet

3 = Blue-violet

4 = Other(describe)



R1

R2

R3

R4

LIGHT SPROUT TIP: INTENSITY OF ANTHOCANIN COLORATION (IF PRESENT)

1 = Absent

2 = Weak

3 = Medium

4 = Strong

5 = Very Strong



R1

R2

R3

R4

LIGHT SPROUT ROOT INITIALS: FREQUENCY

1 = Absent

2 = Some

3 = Abundant



R2

R3

R4

3. PLANT CHARACTERISTICS:

GROWTH HABIT: (See Figure 2)

3 = Erect (>45° with ground)

5 = Semi-erect (30-45° with ground)

7 = Spreading



R1

R2

R3

R4

TYPE:

1 = Stem (foliage open, stems clearly visible)

2 = Intermediate

3 = Leaf (Foliage closed, stems hardly visible)



R1

R2

R3

R4

MATURITY: Days after planting (DAP) at vine senescence



R1

R2

R3

R4

PLANTING DATE:

V

R1

R2

R3

R4

*REGIONAL AREA:

1 = Pacific North West (WA, OR, ID, CO, CA) 4 = Mid-Atlantic Erect (VI, NC, SC, South NJ, FL) 2 = North Central (ND, WI, MI, MN, OH) 5 = South (LA, TX, AZ, NE)

3 = North East (ME, NY, PA, NJ, MD, MA, RI,) 6 = Canada

7 = Europe

8 = England

9 = Latin America

10 = Brazil

11 = Other

R4

V

R1

R2

R3

MATURITY CLASS:

1 = Very Early (<100 DAP) 2 = Early (100-110 DAP) 3 = Mid-season (111-120 DAP) 4 = Late (121-130 DAP) 5 = Very Late (>130 DAP).



R1

R2

R3

4.	STEM CHARACTERISTICS:	Measure at early	v first bloom
╼.	OTEM OTIMICACTERIORIS.	Micasarc at carr	y 1113t biooi11

* STEM ANTHOCYANIN COLORATION:

1 = Absent 3= Weak 5 = Medium 7 = Strong 9 = Very Strong



R1

R2

R3

R4

STEM WINGS: (See Figure 3)

1 = Absent 3 = Weak 5 = Medium 7 = Strong 9 = Very Strong



R1

R2

R3

R4

5. LEAF CHARACTERISTICS:

LEAF COLOR: (Observe fully developed leaves located on middle 1/3 of plant)

1 = Yellowing-green 2 = Olive-green 3 = Medium Green 4 = Dark Green 5 = Grey-green 6 = Other



R1

R2

R3

R4

LEAF COLOR CHART VALUE: Royal Horticulture Society Color Chart or Munsell Color Chart (Observe fully developed leaves located on middle 1/3 of plant and circle the appropriate color chart)



R1

R2

R3

R4

LEAF PUBESCENCE DENSITY:

1 = Absent 2 = Sparse 3 = Medium 4 = Thick 5 = Heavy



R1





R4

LEAF PUBESCENCE LENGTH:

1 = None 2 = Short 3 = Medium 4 = Long 5 = Very Long



R1

R2

R3

R4

(Note Descriptor #15 can be used to describe the type and length of the glandular trichomes observed.)

* LEAF SILHOUETTE: (See Figure 4)

1 = Closed 3 = Medium 5 = Open



R1

R2

R3

R4

PETIOLES ANTHOCYANIN COLORATION:

1 = Absent 3 = Weak 5 = Medium 7 = Strong 9 = Very Strong



R1

R2

R3

R4

LEAF STIPULES SIZE: (Se Figure 5)

1 = Absent 3 = Small 5 = Medium 7 = Large



R1

R2

R3

R4

TERMINAL LEAFLET SHAPE (See Figures 6 and 7)

1 = Narrowly ovate 2 = Medium Ovate 3 = Broadly Ovate 4 = Lanceolate 5 = Elliptical 6 = Obovate 7 = Oblong 8 = Other ______



R1

R2

R3

5. LEAF CHARACTERISTICS: (continued)

TERMINAL LEAFLET TIP SHAPE: (See Figures 6 and 8) 2 = Cuspidate 3 = Acuminate4 = Obtuse5 = Other1 = AcuteR1R2 R3 R4 * TERMINAL LEAFLET BASE SHAPE: (See Figure 9) 3 = Obtuse5 = Truncate 7 = Other4 = Cordate 6 = Lobed1 = Cuneate 2 = Acute**R**1 R2 R3 R4 **TERMINAL LEAFLET MARGIN WAVINESS:** 2 = Slight 3 = Weak 4 = Medium5 = StrongR2 R3 R4 R1NUMBER OF PRIMARY LEAFLET PAIRS: (See Figure 6) AVERAGE: R4 **R3** R1 R2 V RANGE: R4 V R1 R2 **R**3 to to to to to PRIMARY LEAFLET TIP SHAPE: (See Figures 6 and 8) 1 = Acute2 = Cuspidate 3 = Acuminate 4 = Obtuse5 = OtherR2 R3 R1 R4 PRIMARY LEAFLET SIZE: 1 = Very Small 2 = Small3 = Medium4 = Large 5 = Very Large **R**3 R4 **R**1 **R**2 **PRIMARY LEAFLET SHAPE**: (See Figures 6 and 7) 1 = Narrowly ovate 2 = Medium ovate 3 = Broadly ovate 4 = Lanceolate 5 = Elliptical 6 = Ovate 7 = Oblong $8 = Other_$ **R**1 R2 R3 R4 PRIMARY LEAFLET BASE SHAPE: (See Figures 6 and 9) 3 = Obtuse 5 = Truncate 1 = Cuneate 2 = Acute4 = Cordate 6 = Lobed $7 = Other_$ **R**1 R2 R3 R4 NUMBER OF SECONDARY AND TERTIARY LEAFLET PAIRS: (See Figure 6) AVERAGE: R2 R1R3 R4 RANGE:

R2

to

R3

to

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to

R1

to

to

5. LEAF CHARACTERISTICS: (continued)

NUMBER OF INFLORESCENCE/PLANT: AVERAGE: R2 **R3 R**1 R4 RANGE: V **R**1 **R3** R4 to R2 to to to to NUMBER OF FLORETS/INFLORESCENCE: AVERAGE: R1 R2 R3 R4 RANGE: V to **R**1 **R**3 R4 R2 to to to to * COROLLA INNER SURFACE COLOR CHART VALUE: Royal Horticulture Society Color Chart or Munsell Color Chart (Measure predominant color of newly open flower and circle the appropriate color chart) R3 R4 **R**1 R2 * COROLLA OUTER SURFACE COLOR CHART VALUE: Royal Horticulture Society Color Chart or Munsell Color Chart (Measure predominant color of newly open flower and circle the appropriate color chart)

* COROLLA INNER SURFACE COLOR: (Measure predominant color of newly open flower, if flowers are bi-color please use the ratio codes)

1 = White 2 = Red-violet 3 = Blue-violet 4 = Cream 5 = Red-purple 6 = Blue 7 = Pink 8 = Pink-white 9 = Purple 10 = Violet

11 = Purple-violet 13 = Violet-White 1:1 14 = Violet-White 1:3 15 = Violet-White 3:1 16 = Violet-White Halo 17 = Pink-White 1:1 18 =

Pink-White 1:3 19 = Pink-White 3:1 20 = Pink-White Halo 21 = RedViolet-White 1:1 22 = RedViolet-White 1:3 23 = RedViolet-White 3:1

24 = RedViolet-White Halo 25 = BlueViolet-White 1:1 26 = BlueViolet-White 1:3 27 = BlueViolet-White 3:1 28 = BlueViolet-White Halo

12 = Other

R2





R1







R4

R3

COROLLA SHAPE: (See Figure 10)

1 = Very rotate 2 = Rotate 3 = Pentagonal

4 = Semi-stellate 5 = Stellate









R4

6. INFLORESCENCE CHARACTERISTICS:

CALYX ANTHOCYANIN COLORATION:

1 = Absent 3 = Weak 5 = Medium 7 = Strong 9 = Very strong





R2	



R4

ANTHER COLOR CHART VALUE: Royal Horticulture Society Color Chart or Munsel Color Chart (Measure when newly opened flower is fully expanded and circle the appropriate color chart)

4 = Loose



R1

R2



R4

ANTHER SHAPE: (See Figure 11)

1 = Broad cone 2 = Narrow cone 3 = Pear-shaped cone

R1

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R3

5 = Other

6. INFLORESCENCE CHARACTERISTICS: (continued) **POLLEN PRODUCTION:** 1 = None3 = Some5 = AbundantR1R2 **R**3 R4 STIGMA SHAPE: (See Figure 12) 2 = Clavate 3 Bi-lobed 1 = Capitate **R3** R2 R1 R4 STIGMA COLOR CHART VALUE: Royal Horticulture Society Color Chart or Munsel Color Chart (Circle the appropriate color chart) **R**3 R4 **R**1 **R**2 Unofficial Copy BERRY PRODUCTION: (Under field conditions) 7 = Heavy 9 = Very Heavy 1 = Absent3 = Low5 = Moderate R2 R1 R3 **R**4 7. TUBER CHARACTERISTICS: * PREDOMINANT SKIN COLOR: 1 = White 2 = Light Yellow 3 = Yellow4 = Buff5 = Tan6 = Brown7 = Pink8 = Red9 = Purplish-red 10 = Purple 11 = Dark purple-black 12 = OtherR1R2 **R3** R4 PREDOMINANT SKIN COLOR CHART VALUE: Royal Horticulture Society Color Chart or Munsell Color Chart (Circle the appropriate color chart) R3 R4 **R**1 R2 SECONDARY SKIN COLOR: 1 = Absent2 = Present (please describe) **R3** R4 **R**1 R2 SECONDARY SKIN COLOR CHART VALUE: Royal Horticulture Society Color Chart or Munsell Color Chart (Circle the appropriate color) **R3 R**4 R1 R2 SECONDARY SKIN COLOR DISTRIBUTION: (See Figure 13) 3 = Splashed 5 = Spectacled 7 = Other1 = Eyes 2 = Eyebrows4 = Scattered 6 = Stippled R1R2 R3 R4

2 = Rough (flaky)

R1

3 = Netled

4 = Russetted

R2

5 = Heavily russetted

R3

6 = Other

R4

SKIN TEXTURE:

1 = Smooth

7. TUBER CHARACTERISTICS: (continued)

* TUBER SHAPE : (See Figure 14) 1 = Compressed 2 = Round 3 = Oval	4 = Oblong 5 = Lo	ong 6 = Other _		
V R1	R2	R3	R4	
TUBER THICKNESS: 1 = Round 2 = Medium thick 3 = Slight	ntly flattened 4 = Flat	ttened 5 = Othe	r	
V R1	R2	R3	R4	
TUBER LENGTH (mm): AVERAGE:				
V R1	R2	R3	R4	
RANGE:				
V to R1	to R2	to	R3 to	R4 to
STANDARD DEVIATION:				
V R1	R2		R3	R4
AVERAGE WEIGHT OF SAMPLE TAKEN:			1	
V R1	R2		R3	R4
TUBER WIDTH (mm) AVERAGE:				
V R1	R2	R3	R4	
RANGE:				
V to R1	to R2	to	R3 to	R4 to
STANDARD DEVIATION:				
V R1	R2		R3	R4
,				

AVERAGE WEIGHT OF SAMPLE TAKEN (g):

V		R1		R2		R3		R4	
---	--	----	--	----	--	----	--	----	--

7. TUBER CHARACTERISTICS: (continued)

TUBER THICKNESS (mm):

AVERAGE:



R1

R2

R3

R4

RANGE:



R1 to

R2 to

R3 to

R4 to

STANDARD DEVIATION:



R1

R2

R3

R4

AVERAGE WEIGHT OF SAMPLE TAKEN (g):



R1

R2

R3

R4

TUBER EYE DEPTH:

1 = Protruding

3 = Shallow

5 = Intermediate

7 = Deep

9 = Very deep



R1

R2

R3

R4

TUBER LATERAL EYES:

1 = Protruding

3 = Shallow

5 = Intermediate

e 7 = Deep

9 = Very deep



R1

R2

R3

R4

NUMBER EYE/TUBER:

AVERAGE:



R1

R2

R3

R4

RANGE:

V to)
------	---

R1

R2 to

R3 to

R4 to

DISTRIBUTION OF TUBER EYES:

1 = Predominantly apical

2 = Evenly distributed

to



R1

R2

R3

R4

PROMINENCE OF TUBER EYEBROWS:

1= Absent

2 = Slight prominence

3 = Medium prominence

4 = Very prominent

5 = Other _____

V

R1

R2

R3

7. TUBER CHARACTERISTICS: (continued)

PREDOMINANT TUBER FLESH COLOR

1 = White 2 = Light Yellow 3 = Yellow4 = Buff5 = Tan6 = Brown7 = Pink8 = Red9 = Purplish-red 10 = Purple 11 = Dark purple-black 12 = Other

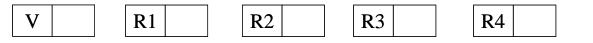
R3 R4 V **R**1 R2

PRIMARY TUBER FLESH COLOR CHART VALUE: Royal Horticulture Society Color Chart or Munsell Color Chart (Circle the appropriate color chart)

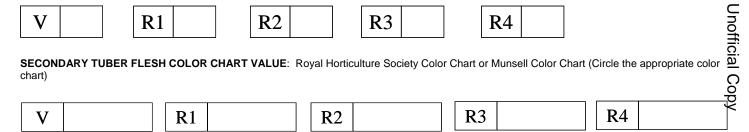
R3 R4 V **R**1 R2

SECONDARY TUBER FLESH COLOR:

1 = Absent2 = Present, please describe:



SECONDARY TUBER FLESH COLOR CHART VALUE: Royal Horticulture Society Color Chart or Munsell Color Chart (Circle the appropriate color chart)



NUMBER OF TUBERS/PLANT:

2 = Medium (8-15)3 = High (>15)1 = Low (< 8)



8. DISEASES CHARACTERISTICS:

DISEASES REACTION: 0 = Not Tested 1 = Highly Resistant 2 = Resistant Few Symptoms 3 = Resistance Few Lessions in Number and Size 4 = Moderately Resistance 5 = Intermedia Susceptible 6 = Moderate Susceptible

7 = Susceptible 9 = Highly Susceptible

LATE BLIGHT: (Phytophthora)



R2

R3

R4

EARLY BLIGHT: (Alternaria)



R1

R2

R3

R4

SOFT ROT (Erwinia)



R1

R2

R3

R4

COMMON SCAB (Streptomyces)



R1

R2

R3

R4

POWDERY SCAB (Spongospora)



R1

R2

R3

R4

DRY ROT (Fusarium)



R1

R2

R3

R4

POTATO LEAF ROLL VIRUS (PLRV)



R1

R2

R3

8. DISEASES CHARACTERISTICS: (continued)

POTATO VIRUS X (PVX)



R1

R2

R3

R4

POTATO VIRUS Y (PVY)



R1

R2

R3

R4

POTATO VIRUS M (PVM)



R1

R2

R3

R4

POTATO VIRUS A (PVA)



R1

R2

R3

R4

GOLDEN NEMATODE (Globodera)



R1

R2

R3

R4

ROOT – KNOT NEMATODE (Meloidogyne)



R1

R2

R3

R4

OTHER DISEASE



R1

R2

R3

R4

PHYSIOLOGICAL DISORDER

1 = Malformed shape6 = Blackheart

2 = Tuber cracking 7 = Internal sprouting

3 = Feathering ng 8 = Other

4 = Hollow heart

5 = Internal necrosis

V

R1

R2

R3

R4

9. PESTS CHARACTERISTICS:

PEST REACTION: 0 = Not Tested 1 = Highly Resistant 2 = Resistant Few Symptoms 3 = Resistance Few Lessions in Number and Size

4 = Moderately Resistance 5 = Intermedia Susceptible 6 = Moderate Susceptible

7 = Susceptible 9 = Highly Susceptible

COLORADO POTATO BEETLE (CPB) (Leptinotarsa)



R1

R2

R3

R4

GREEN PEACH APHID (Myzus)



R1

R2

R3

R4

OTHER:



R1

R2

R3

R4

OTHER:



R1

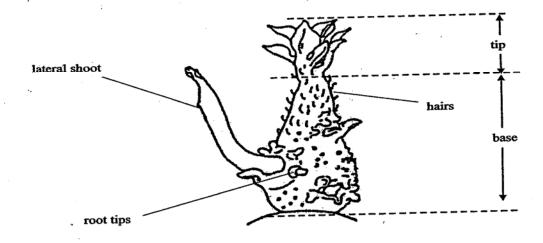
R2

R3

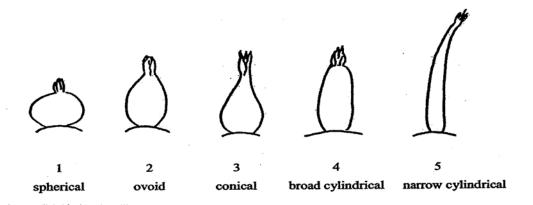
10. G	ENE TRAITS:					1
	INSERTION OF	GENES : 1 = YES 2 =	: NO			1
	IF YES, describe	e the gene(s) introduced of	or attach information:			
11. Q	JALITY CHARACTI	ERISTICS:				
	CHIEF MARKET	Γ :				
		VITY (wt. air/wt. air – wt. v				
	1 = <1.060	2 = 1.060-1.069 3 =	1.070-1.079 4 = 1	1.080-1.089 5 = >1.090)	į
	V	R1	R2	R3	R4	
	TOTAL GLYCO	ALKALOID CONTENT (r	ng./100 g. fresh tuber)			
	V	R1	R2	R3	R4	<u> </u>
						Ç
OTHE baking	R QUALITY CHARA , boiling, after-cookii	ACTERISTICS: Describe ng darkening). Please at	any other quality chara tach data and correspo	acteristics that may aid in i inding protocol.	dentification, (e.g., chip-processing	ي, french fry processing, ي ر
	_					
12. C	HEMICAL IDENTIFI	CATION:				
Descri	be chemical traits of	the candidate variety tha	t aid in its identification	(e.g., protien or DSN elec	etrophoresis). Please attach data a	and the corresponding
protoc		•			,	, ,
42 FI	NGER PRINTING M	IADVEDE.				
13. FI						
	ISOZYMES 1:					
	IF YES, attach ir	nformation				
14. DI	NA PROFILE: 1 =	YES 2 = NO				
	IF YES, attach ir					
	ii 120, attaorri	Homaton				
15. A	ODDITIONAL COM	MENTS AND CHARACTI	ERISTICS:			
Include	e any additional desc	criptors that would be use	ful in distringuishing the	e candidate variety.		

Figure 1: Light sprout

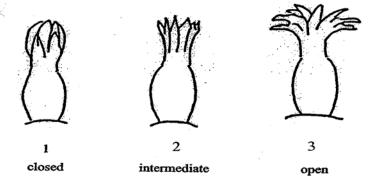
Light sprout dissection



Light sprout shape



Light sprout tip habit

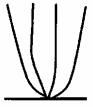


The characteristic should be observed after about 10 weeks to obtain a good differentiation in the collection.

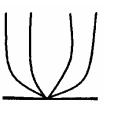
Figure 2: Growth Habit







Semi Erect



Spreading

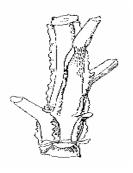
Figure 3: Stem Wings



Weak



Medium



Strong

Figure 4: Leaf Sillhouette



Closed

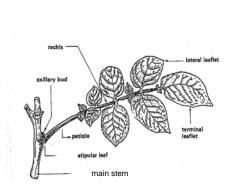


Medium

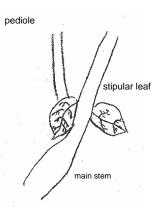


Open

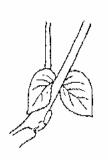
Figure 5: Leaf Stipules



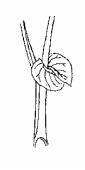
General structures



Small stipular leaf



Medium stipular leaf



Large stipular leaf

Figure 6: Leaf Dissection

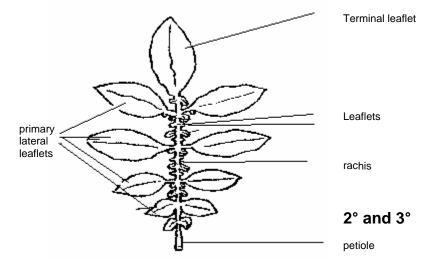


Figure 7: Terminal Leaflet Shape/Primary Leaflet Shape

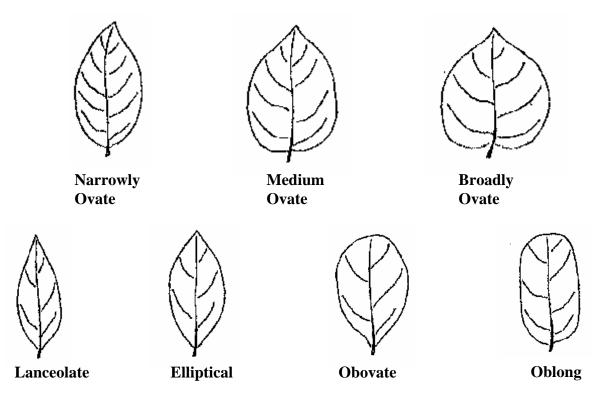


Figure 8: Terminal Leaflet Shape of Tip/Primary Leaflet Shape of Tip

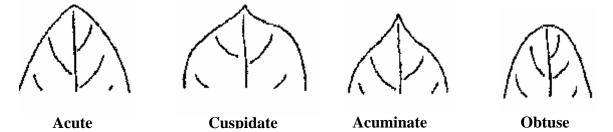


Figure 9: Terminal Leaflet Shape of Base/Primary Leafelet Shape of Base

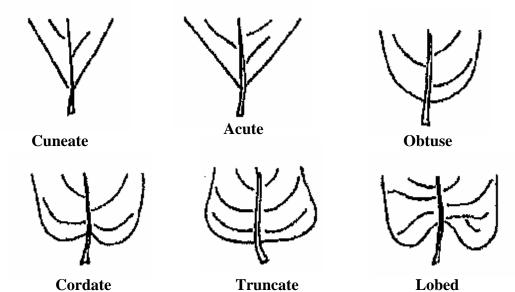


Figure 10: Corolla Shape

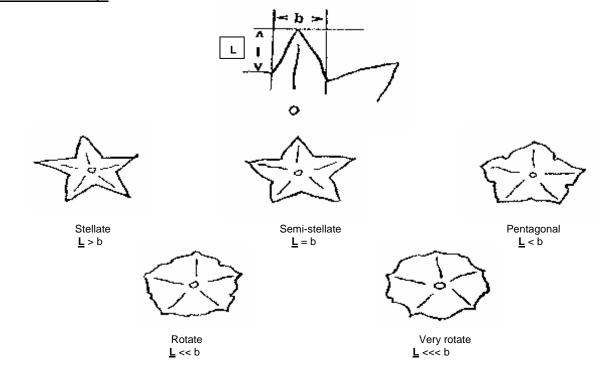


Figure 11: Anther Shape



Broad cone



Narrow cone



Pear-shape cone



Loose

Figure 12: Stigma Shape

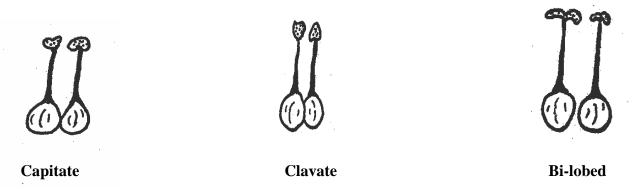


Figure 13: Distribution of Secondary Skin Tuber Color

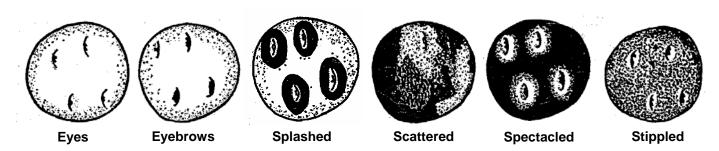
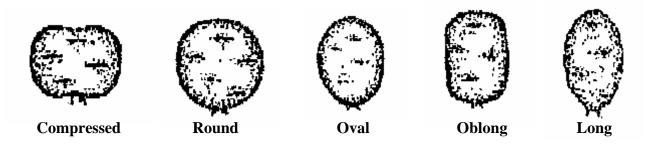


Figure 14: Tuber Shape



References:

Huaman, Z. 1986. Systematic botany and morphology of the potato. Technical information Bulletin 6. International Potato Center, Lima, Peru.

Huaman, Z., Williams, J.T., Salhuana, W. and Vincent, L. Descriptors for the cultivated potato and the maintenance and distribution of germplasm collections. 1977. International Board for Plant Genetic Resources. Rome, Italy.

Potato (Solanum tuberosum L.) Guidelines for the conduct of tests for distinctness, uniformity and stability. International union for the protection of new varieties of plants (UPOV). 2004-03-31.

GENERAL INSTRUCTIONS: To be effectively filed with the Plant Variety Protection Office (PVPO), ALL of the following items must be received in the PVPO: (1) Completed application form signed by the owner: (2) completed exhibits A. B. C, E, F; (3) for a tuber reproduced variety, verification that a viable (in the sense that it will reproduce an entire plant) tissue culture will be deposited and maintained in an approved public repository; and (4) payment by credit card or check drawn on a U.S. bank for \$4,382 (\$518 filing fee and \$3,864 examination fee), payable to "Treasurer of the United States" (See Section 97.6 of the Regulations and Rules of Practice). NEW: With the application for a seed reproduced variety or by direct deposit soon after filing, the applicant must provide at least 3,000 viable untreated seeds of the variety per se, and for a hybrid variety at least 3,000 untreated seeds of each line necessary to reproduce the variety. Partial applications will be held in the PVPO for not more than 90 days, then returned to the applicant as un-filed. Mail application and other requirements to Plant Variety Protection Office, AMS, USDA, Room 401, NAL Building, 10301 Baltimore Avenue, Beltsville, MD 20705-2351. Retain one copy for your files. All items on the face of the application are self explanatory unless noted below. Corrections on the application form and exhibits must be initialed and dated. DO NOT use masking materials to make corrections. If a certificate is allowed, you will be requested to send a payment by credit card or check payable to "Treasurer of the United States" in the amount of \$768 for issuance of the certificate. Certificates will be issued to owner, not licensee or agent.

NOTES: It is the responsibility of the applicant/owner to keep the PVPO informed of any changes of address or change of ownership or assignment or owner's representative during the life of the application/certificate. The fees for filling a change of address; owner's representative; ownership or assignment; or any modification of owner's name is specified in Section 97.175 of the regulations. (See Section 101 of the Act, and Sections 97.130, 97.131, 97.175(h) of the Regulations and Rules of Practice.)

Plant Variety Protection Office

FAX: (301) 504-5291 Telephone: (301) 504-5518

General E-mail: PVPOmail@usda.gov

Homepage: http://www.ams.usda.gov/science/pvpo/PVPindex.htm

SPECIFIC INSTRUCTIONS:

To avoid conflict with other variety names in use, the applicant must check the appropriate recognized authority and provide evidence that the permanent name of the application variety (even if it is a parental, inbred line) has been cleared by the appropriate recognized authority before the Certificate of Protection is issued. For example, for agricultural and vegetable crops, contact: U.S. Department of Agriculture, Agricultural Marketing Service, Livestock and Seed Programs, Seed Regulatory and Testing Branch, 801 Summit Crossing Place, Suite C, Gastonia, North Carolina 28054-2193 Telephone: (704) 810-8870. http://www.ams.usda.gov/lsg/seed.htm.

ITEM

19a. Give:

- (1) the genealogy, including public and commercial varieties, lines, or clones used, and the breeding method;
- (2) the details of subsequent stages of selection and multiplication;
- (3) evidence of uniformity and stability; and
- (4) the type and frequency of variants during reproduction and multiplication and state how these variants may be identified
- 19b. Give a summary of the variety's distinctness. Clearly state how this application variety may be distinguished from all other varieties in the same crop. If the new variety is most similar to one variety or a group of related varieties:

 - identify these varieties and state all differences objectively;
 attach replicated statistical data for characters expressed numerically and demonstrate that these are clear differences; and
 - (3) submit, if helpful, seed and plant specimens or photographs (prints) of seed and plant comparisons which clearly indicate distinctness.

19c. Exhibit C forms are available from the PVPO Office for most crops; specify crop kind. Fill in Exhibit C (Objective Description of Variety) form as completely as possible to describe your variety.

- 19d. Optional additional characteristics and/or photographs. Describe any additional characteristics that cannot be accurately conveyed in Exhibit C. Use comparative varieties as is necessary to reveal more accurately the characteristics that are difficult to describe, such as plant habit, plant color, disease resistance, etc.
- 19e. Section 52(5) of the Act requires applicants to furnish a statement of the basis of the applicant's ownership. An Exhibit E form is available from the PVPO.
- If "Yes" is specified (seed of this variety be sold by variety name only, as a class of certified seed), the applicant MAY NOT reverse this affirmative decision after the variety has been sold and so labeled, the decision published, or the certificate issued. However, if "No" has been specified, the applicant may change the choice. (See Regulations and Rules of Practice, Section 97.103).
- 23. See Sections 41, 42, and 43 of the Act and Section 97.5 of the regulations for eligibility requirements.
- 24. See Section 55 of the Act for instructions on claiming the benefit of an earlier filing date.
- 22. CONTINUED FROM FRONT (Please provide a statement as to the limitation and sequence of generations that may be certified.)
- 23. CONTINUED FROM FRONT (Please provide the date of first sale, disposition, transfer, or use for each country and the circumstances, if the variety (including any harvested material) or a hybrid produced from this variety has been sold, disposed of, transferred, or used in the U.S. or other countries.)

Application filed within one year of release date.

24. CONTINUED FROM FRONT (Please give the country, date of filing or issuance, and assigned reference number, if the variety or any component of the variety is protected by intellectual property right (Plant Breeder's Right or Patent).)

According to the Paperwork Reduction Act of 1995, an agency may not conduct or sponsor, and a person is not required to respond to a collection of information unless it displays a valid OMB control number for this information collection is 0581-0055. The time required to complete this information collection is estimated to average 1.4 hours per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information.

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